



UNIVERSIDAD COMPLUTENSE DE MADRID
Facultad de Veterinaria
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STANDARD PROCEDURE OPERATION ELISA FOR SEROLOGICAL DIAGNOSIS FOR AFRICAN SWINE FEVER

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1. MATERIALS AND REAGENTS

Watch video (Indirect ELISA-OIE)

- Single channel pipettes 1-10 μ l
 - Single channel pipettes 10-100 μ l
 - Single channel pipettes 10-200 μ l
 - Single channel pipettes 200-1000 μ l
 - Adsorbent paper
 - Aluminium foil
 - Multichannel pipette 5-50 μ l
 - Multichannel pipette 50-300 μ l
 - NUNC-Polysorp microtiter plate (ref: 469957.NUNC)
 - Chamber 37°C
 - Analytical Balance
 - Distilled water
 - Minincubationn trays (ref:170-3902.BIORAD)
 - Ph meter
 - Tubes shaker or vortex mixer
 - Reagent reservoir Polystyrene 50ml (ref: 4870.COSTAR)
 - Steril plastic tubes (10ml,50ml)
 - Spectrophotometre UV/VIS with filter 620 nm annexed to a computer program to register result.
 - Table centrifuge
 - Latex or nitrile gloves
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- **Ag**: Antigen supplied by ASF reference laboratory in lyophilized vials of 0,5 ml, 1ml or 2ml. Store once reconstituted aliquot and freze at -20°C. Expiry date: 18 months.
 - **PC**: reference positive control serum supplied by ASF reference laboratory in lyophilized vials of 0,5 ml, 1ml or 2ml. Store once reconstituted aliquot and freeze at -20°C. Expiry date: 18 months.
 - **LC**: reference limit control control serum supplied by ASF reference laboratory in lyophilized vials of 0,5 ml, 1ml or 2ml. Store once reconstituted aliquot and freeze at -20°C. Expiry date: 18 months.
 - **NC**: reference negative control serum supplied by ASF reference laboratory in lyophilized vials of 0,5 ml, 1ml or 2ml. Store once reconstituted aliquot and freeze at -20°C. Expiry date: 18 months.
 - **CONJUGATE**: Proteina A peroxidase 1mg/ml



• **CARBONATE/BICARBONATE BUFFER 0,05M (pH 9,6):**

1,59 gr ----- Na_2CO_3 [Merck 1.06392]
2,93 gr ----- NaHCO_3 [Merck 10 6329]
1 L. ----- Distilled water

Store at room temperature. Check the pH(9,6) before use:

↑ pH: Sodium bicarbonate

↓ pH: Sodium carbonate

• **HYDROGEN PEROXIDASE (H₂O₂)**

To use at 30 %.

• **WASHING SOLUTION; PBS-TWEEN BUFFER Ph 7,2:**

ClNa [Merck 1.06404]----- 8 gr
ClK [Merck 1.04873]----- 0,2 gr
 $\text{PO}_4\text{H}_2\text{K}$ [Merck 1.06586]----- 0,2 gr
 PO_4HNa_2 [Merck 1.04936]----- 1,15gr
Tween-20 [Merck 8.22184]----- 0,5 ml
 H_2O distilled ----- 1000 ml

Store at room temperature. Check the pH before use.

• **SUBSTRATE SOLUTION:**

❖ **DMAB -3- Dimethylaminobenzoic acid:**

Dissolve 13,315 gr DMAB in 900 ml of *phosphate buffer 0,1 M pH 7*

-Phosphate buffer 0,1 M pH 7:

5,3 g ----- $\text{PO}_4\text{H}_2\text{K}$
8,65 g ----- PO_4HNa_2
1 l. ----- H_2O distilled

Mix during 1 hour at room temperature, adjust pH to 7 with NaOH 5M.
Adjust the final volume to 1 L. Filter and prepare aliquots of 10ml and
5ml. *Storage at -20°C in darkness.*



❖ MBTH -3- Methyl-2-benthiiazolinone hydrazone hydrochloride monohidrate:

Dissolve 0,3646 gr MBTH in 900 ml of *phosphate buffer 0,1 M pH 7*. Mix during 1 hour at room temperature, adjust pH to 6,25 with concentrate HCl. After that, adjust the final volume to 1 litre. Filter and prepare aliquots of 10ml and 5ml. *Storage at -20°C in darkness.*

• STOP SOLUTION (Sulphuric acid 3N):

Sulphuric Acid ----- 16,1 ml (in 200 ml distilled water).
Storage at room temperature.

2. METHODOLOGY:

Watch video (Indirect ELISA-OIE)

- ❖ NOTE: Before the ELISA assay, plates must be sensitised with the antigen. In this video plates were already sensitised.

Sensitisation of microtiter plates with antigen:

- Dilute the soluble antigen in carbonate/bicarbonate buffer pH 9,6. at the recommended concentration.
e.g.: (Ag 1/1600): 6,25µl of Ag + 9,99 ml carbonate/bicarbonate buffer pH 9,6
- Add 100 µl per well of a NUNC-Polysorp microtiter plate.
- Incubate at 4 °C for 18h (overnight)

The sensitised and dry plates can be stored at 4°C for one day or at -20°C for several months.

2.1 Wash the plates four times with washing buffer. Blot them onto paper towels.

2.2 Dilute test and control sera (1/30) in PBS/Tween-20 solution
- Add 96,6 µl of PBS 0,5 tween-20 to the **NON sensitised** plates



- Label the NON sensitised plate, where we are going to dilute test and control sera.
- Add 3,4 μ l of test and control sera.

2.3 Add 100 μ l of each diluted serum in duplicate to plate wells. A recommended plate design includes duplicate control sera.

2.4 Cover the plate and incubate for 1 h at 37 °C in agitation.

2.5 Wash the plates four times with washing buffer. Then blot them onto paper towels.

2.6 Conjugate preparation (1/5000):

*Add 2 μ l Protein A to 9998 μ l of PBS 0,5 Tween 20
(Volume for one plate)*

2.7 Add 100 μ l of conjugate per well.

2.8 Cover the plate and incubate for 1 h at 37 °C in agitation.

2.9 Wash the plates four times with washing buffer. Then blot them onto paper towels.

2.10 Substrate solution preparation:

*10ml of DMAB + 10 ml of MBTH + 5 μ l H₂O₂ (30%)
(Volume for one plate)*

2.11 Add 200 μ l of substrate solution per well.

2.12 Cover the plate with aluminium foil. Incubate for approximately 5-10 minutes in darkness at room temperature or until observe that Negative Control begin to take colour.

2.13 Stop the reaction by addition of 50 μ l stop solution per well.

2.14 Reading plates. The results can be obtained using a spectrophotometer UV/VIS to read microtiter plates at 620 nm wavelengths.



3. RESULTS

➤ VALIDATION OF THE TEST:

The test could be considered valid when the OD of the PC (Positive Control) is, at least, 4 times higher than the OD of the NC (Negative Control).

$$DO PC \geq 4 DO NC$$

Value of OD PC must be ≥ 1.0

Value of OD NC must be ≤ 0.250

Value of OD LC must be in range of Cut Off, with a value of OD up to 0,7.

➤ CUT OFF CALCULATION:

$$CUT OFF = (OD \text{ negative serum}) + (OD \text{ Positive serum} \times 0,2)$$

- **Negative sera:** OD below the CUT OFF -0,1.
- **Positive sera:** OD higher than CUT OFF + 0,100.
- **Ambiguous sera:** OD between CUT OFF +/- 0,100. They have to be confirmed by IB technique.